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# Yusdy Pan, MSc, PhD, Singapore

- Principal Scientist, Process Development,  
Amgen Singapore Manufacturing



# Harvest process in commercial biologicals manufacturing

Yusdy Pan, MSc, PhD

5 August 2018



# HARVEST PROCESS IN COMMERCIAL BIOLOGICALS MANUFACTURING

**YUSDY PAN PH.D.**

PRINCIPAL SCIENTIST, PROCESS DEVELOPMENT

AMGEN SINGAPORE MANUFACTURING

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# AGENDA

- Small synthetic vs large biologics modalities
- Conventional harvest process in biologics manufacturing
- New harvest technologies for high cell density cell culture process

## Disclaimer:

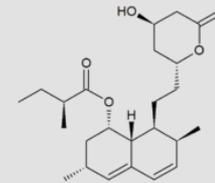
*This presentation do not represent what we do at Amgen, but represent current technologies across industry with examples of vendors for each technology. Amgen does not provide any endorsement for vendors listed in this presentation*

# BIOLOGICS ARE MORE COMPLEX THAN SMALL SYNTHETIC MOLECULE THERAPEUTICS

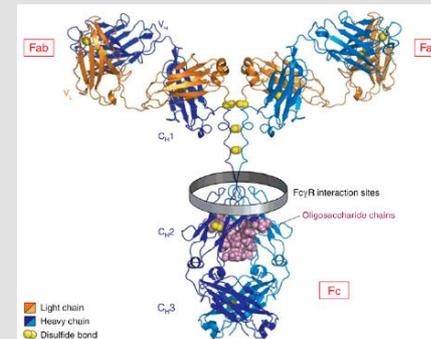


Small Synthetic Molecules	Biologics
Chemical ingredients in simple structures	Proteins produced by living systems
Relatively stable	Variable; sensitive to conditions
Defined structure and easy to characterize	Heterogeneous structures and difficult to characterize

courtesy of D. Fenton



Lovastatin, the first statin to be marketed  
MW: 404.55

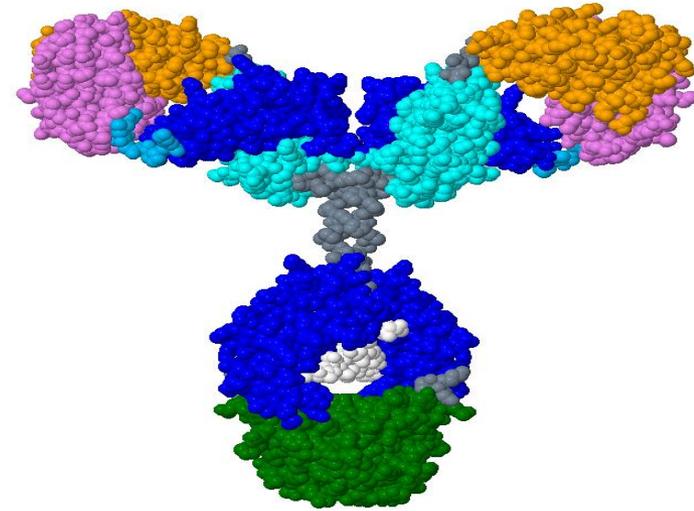


IgG  
MW:  
150,000

**COMPARED TO SYNTHETIC, BIOLOGICS DRUGS ARE MORE SPECIFIC TO TARGET & THEREFORE LESS TOXIC**

# MONOCLONAL ANTIBODIES

- Antibodies are a class of proteins known as immunoglobulins
- Found in blood or other bodily fluids of vertebrates
- Used by the immune system to identify and neutralize foreign organisms and molecules
- The huge diversity of antibodies allows the immune system to recognize an equally wide diversity of antigens



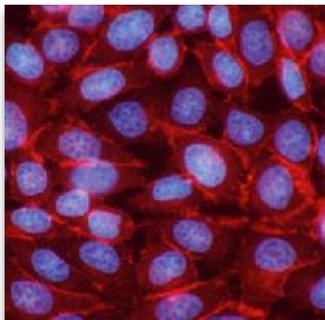
Large, complex molecule  
Molecular Weight: 150 kDa

It has been estimated that humans generate about 10 billion different antibodies, each capable of binding a distinct antigen

# MAMMALIAN VS MICROBIAL EXPRESSION SYSTEMS

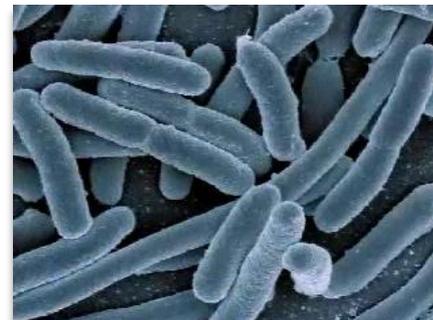
## Mammalian Cells

- **Slower, more complicated cell growth**
- **Similar protein processing to humans (e.g. post translational modification)**
- **Less complicated recovery and purification**

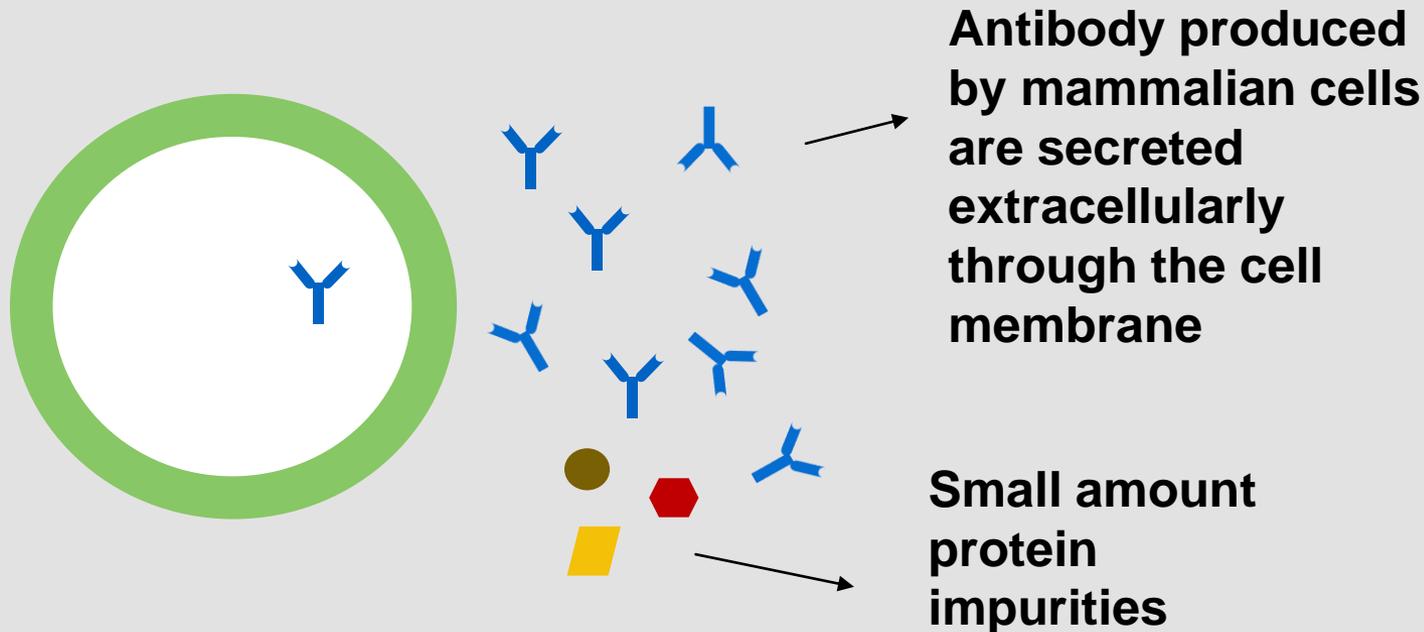


## Microbial Cells

- **Faster, relatively straightforward fermentation**
- **High-yield inclusion bodies**
- **Difficult purification with lower yield**
- **Do not work for all protein therapeutics**



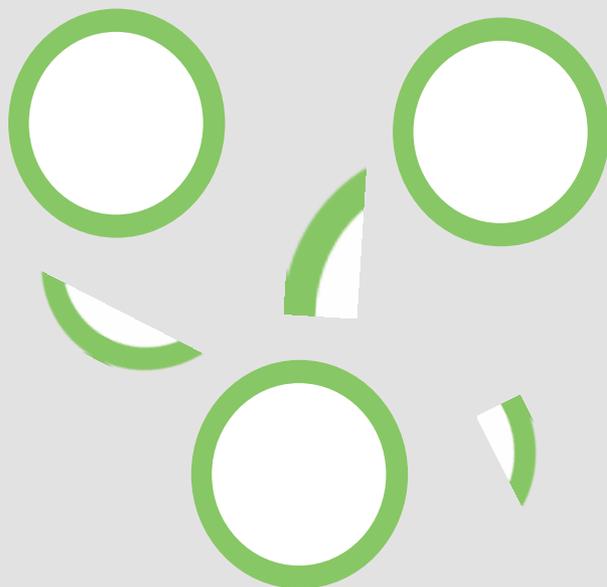
# MAMALLIAN CELL EXPRESSION SYSTEM



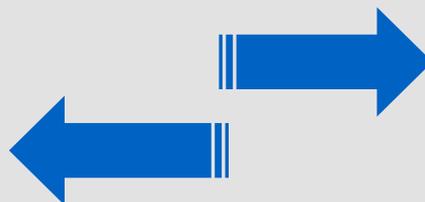
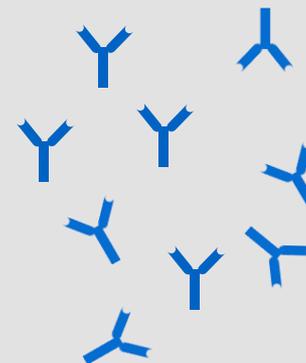
**Antibody recovery process is relatively simpler with mammalian cell expression system and do not require cell disruption**

# HARVEST FOR MAMALLIAN CELL PROCESS

Cells & cell debris



Antibody



**OBJECTIVE OF HARVEST PROCESS IS TO RECOVER THE ANTIBODY PRODUCT FROM CELLS & CELL DEBRIS**

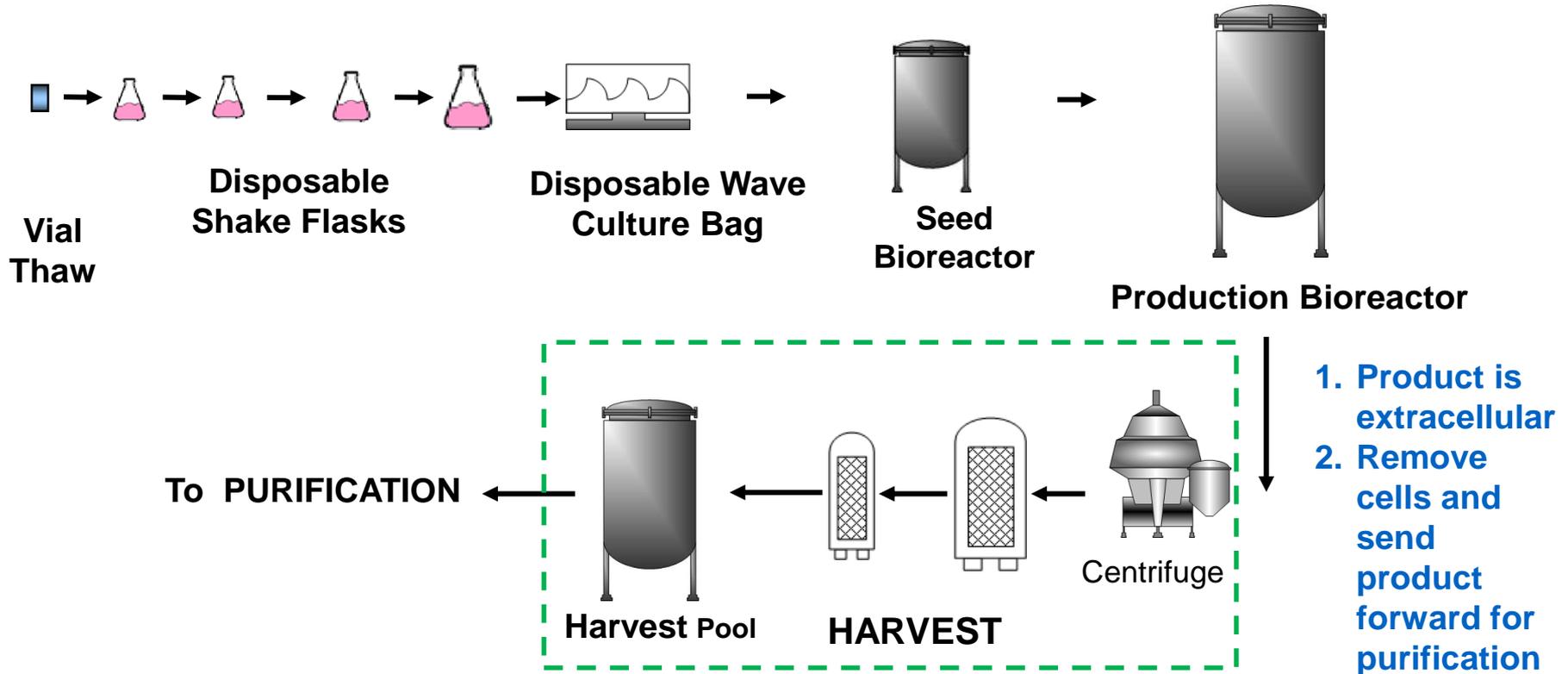
# PRINCIPLE IN CELL SEPARATION TECHNIQUES

- **SIZE** (Cells are bigger than proteins)
  - Filtration
  - Retainment
- **DENSITY** (Cells are heavier than proteins)
  - Centrifugation
  - Sedimentation

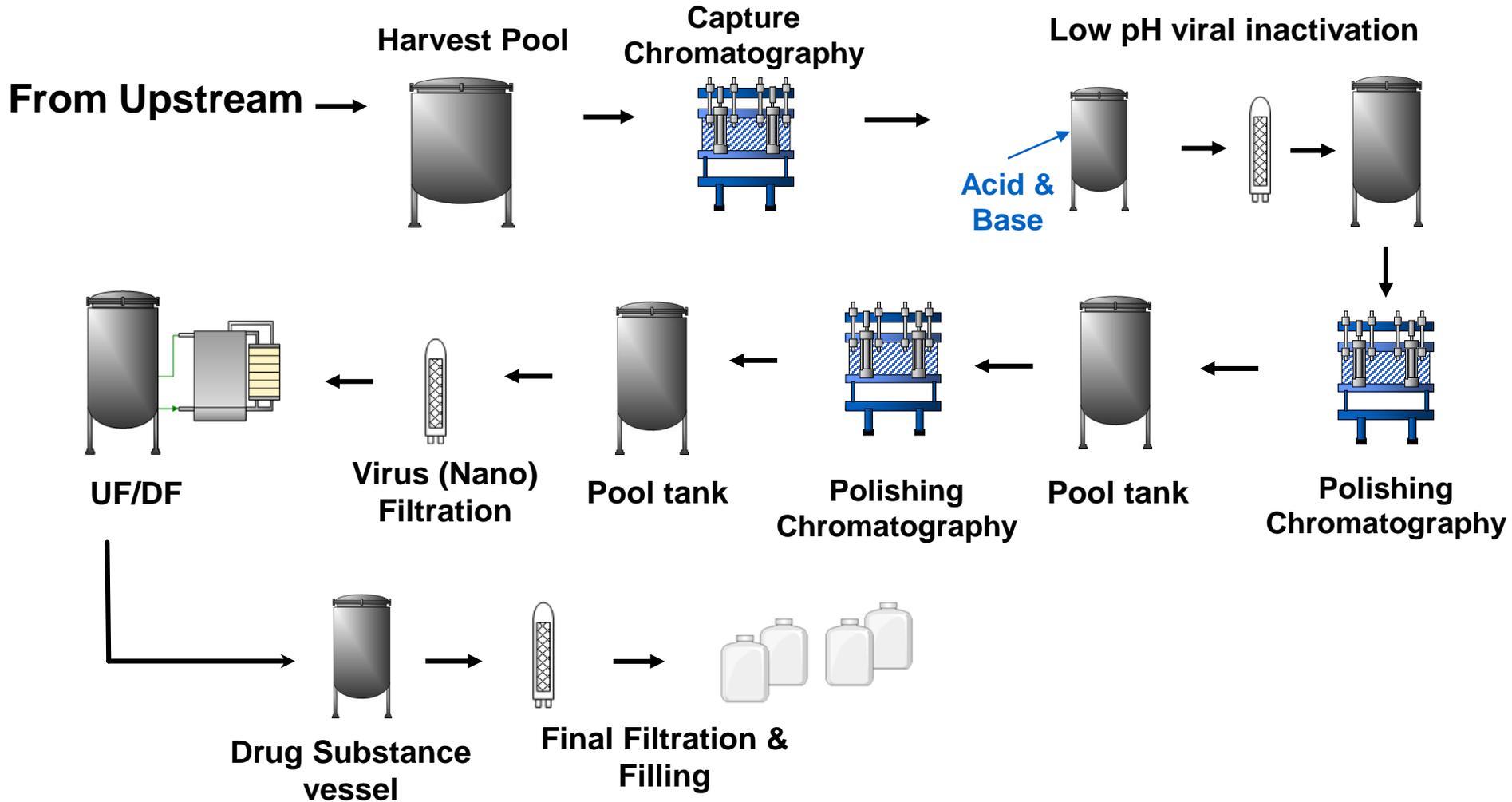


# CONVENTIONAL HARVEST PROCESS IN MAMALLIAN CELL CULTURE PROCESS

# TYPICAL MAMALLIAN CELL CULTURE & HARVEST PROCESS FOR MONOCLONAL ANTIBODY PRODUCTION



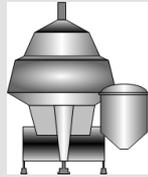
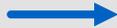
# TYPICAL 3 COLUMN STEP DOWNSTREAM PROCESS



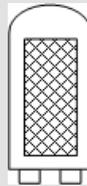
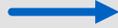
# CONVENTIONAL HARVEST PROCESS IN MAMALLIAN CELL CULTURE PROCESS



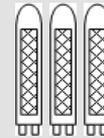
**Bioreactor**  
Last day of production



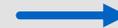
**Centrifugation**  
Remove intact cells & large debris (micron size)



**Depth filtration**  
Remove cell debris ( Micron and sub-micron sizes)



**Membrane filtration**  
Served as an aseptic boundary



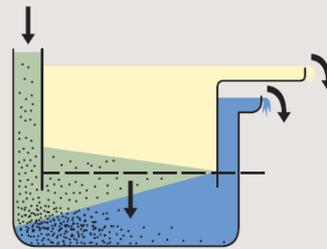
**Harvest Vessel**  
Product solution, free of cell and debris, ready for purification

**Harvest process with combination of centrifugation + filtration.  
Suitable for cell culture process with < ~10 million of cell density**

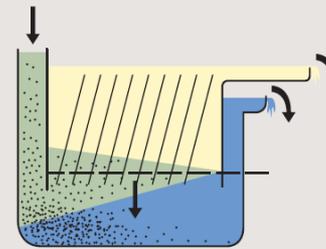
# DISC STACK CENTRIFUGE



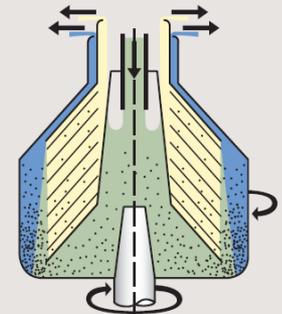
Contain multi disc stacks for efficient separation



Settling tank



Settling tank with discs



Disc stack separator

Mechanism of separation is similar to a settling decanter

Efficiency of separation depends on centrifuge rpm speed, residence time, size and no of disc, density difference between particle and solution

*Pictures courtesy of Alfa Laval. Alfa laval is an example of centrifuge vendors*

# DISC STACK CENTRIFUGE

## Design Parameters:

- Equivalent clarification surface area ( $\Sigma$ )
  - Centrifuge design (no of discs, dimension)
  - Bowl speed (rpm)
- Flow rate (Q)
- Feed material Cell culture viability
  - Cell density (% PCV -> pack cell volume)
  - Cell culture viability
- Solid discharge interval (volumetric, time, turbidity)
  - e.g. A centrifuge has a solid bowl capacity of 10L (target 80% fill). Cell culture feed of 2% PCV at 100LPM. Solid discharge interval/frequency can be set at  $10L \times 80\% / (100LPM \times 2\%) =$

$$4 \text{ min} = 400L \text{ flowthrough}$$

BioProcess International, NOV 2007, 38-50

$$\Sigma = \frac{2\pi}{3g} \times \omega^2 \times N \times \cot \alpha \times (r_o^3 - r_i^3)$$

where

$\Sigma$  = equivalent clarification area of centrifuge (m<sup>2</sup>)

$g$  = acceleration due to gravity (m/s<sup>2</sup>)

$\omega$  = angular bowl velocity (rad/s)

$N$  = number of discs in stack

$\alpha$  = disc half-conical angle (°)

$r_o$  = outer radius of disc (m)

$r_i$  = inner radius of disc (m)

$$d_{\min} = \sqrt{\frac{Q}{\Sigma}} \times \sqrt{\frac{18\eta}{\Delta\rho g}}$$

where

$d_{\min}$  = diameter of minimum particle size that can be separated (m)

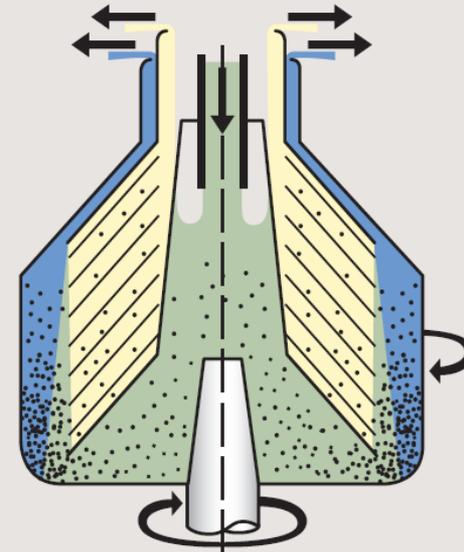
$Q$  = centrifuge feed rate (m<sup>3</sup>/s)

$\Sigma$  = equivalent separation area of centrifuge (m<sup>2</sup>)

$\eta$  = dynamic viscosity of culture medium (kg/m·s)

$\Delta\rho$  = density difference between cells and medium (kg/m<sup>3</sup>)

$g$  = acceleration due to gravity (m/s<sup>2</sup>)



Picture from 'Alfa Laval – disc stack separator technology' brochure

**Q /  $\Sigma$  factor is maintained constant during scale up/down**

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# DEPTH FILTRATION

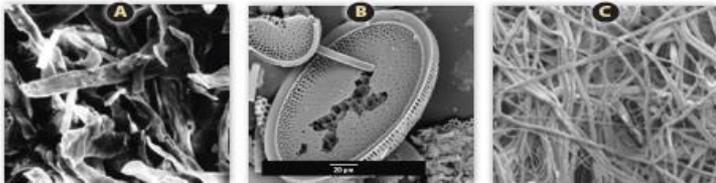
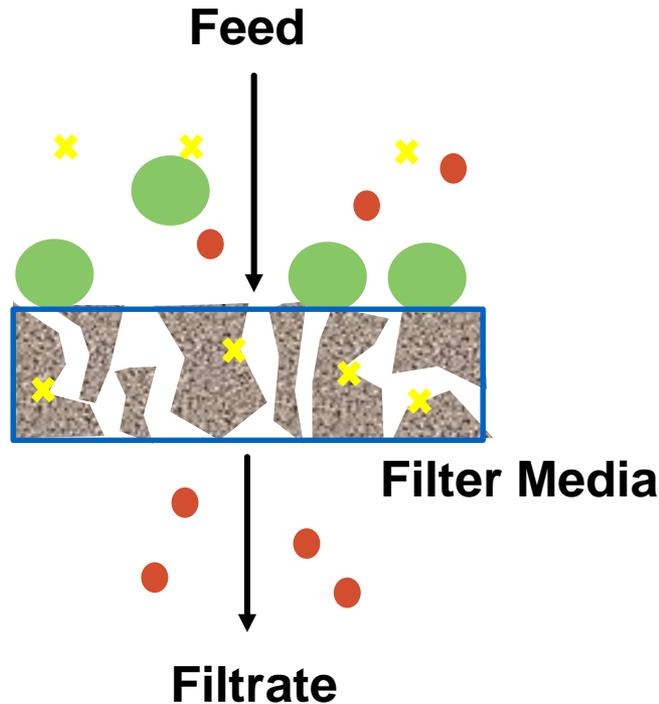


FIGURE 1. Main components of depth filters: (A) cellulose fibers; (B) diatomaceous earth; and (C) polypropylene fibers.

- Depth filters are typically composed of cellulose fibers impregnated with diatomaceous earth and polymeric binding additives
- Mechanism of separation : sieving & adsorption
- Filter media may contain charge species (typically positive charge) which can bind Host Cell Protein (HCP) and DNA
- Pre-use water flush is required to remove extractables & leachables

# DEPTH FILTRATION

POD disposable depth filter system, EXAMPLE from MerckMillipore



Lenticular depth filter system, example from 3M (available in disposable or SS housing format)



Zeta Plus™ Cartridges and Capsule Family

## Unparalleled Scalability

The Pod holder system's modular design makes it easy to configure a system for a specific application and conveniently reconfigure it as process capacity requirements scale-up or down. The flexible, modular format offers scalability from 5 to 12,000 liters or more.



Small scale



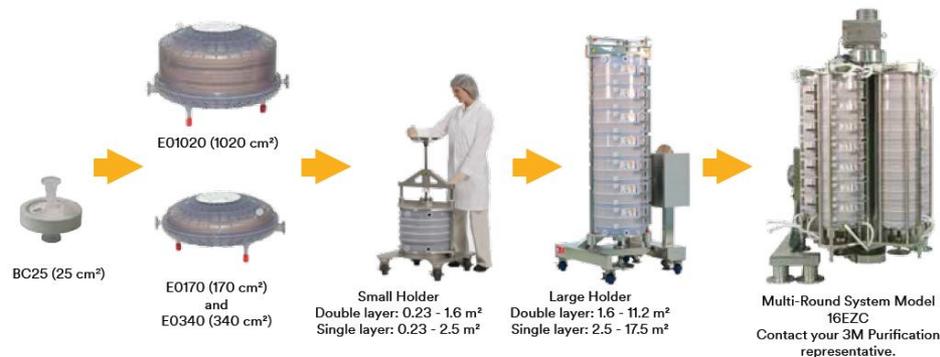
Lab scale



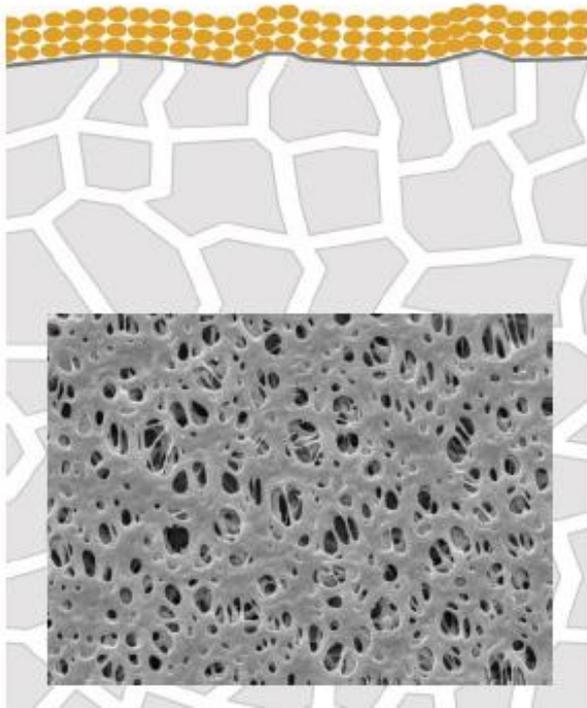
Pilot scale



Process scale

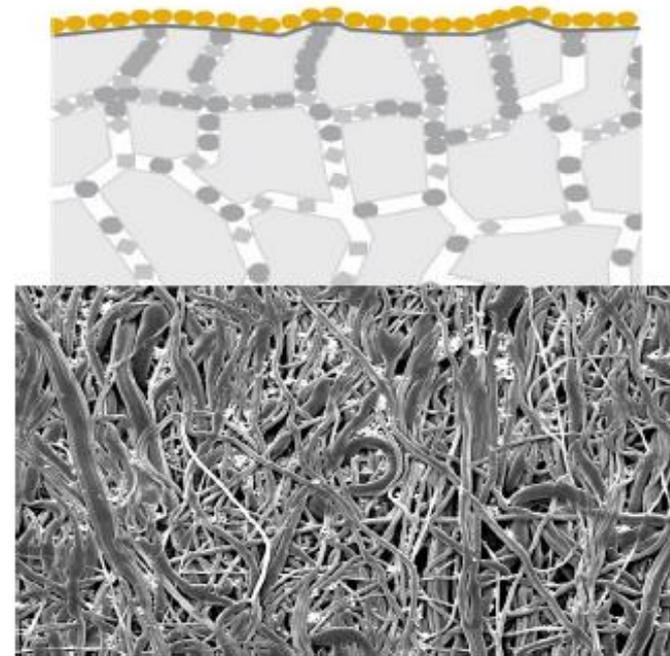


# MEMBRANE FILTER VS DEPTH FILTER



Surface filtration

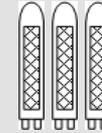
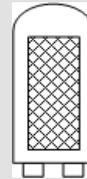
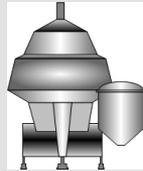
*Membrane Filters*



Depth filtration

*Depth Filter Sheets / Modules*

# COMMON PROCESS MONITORING IN CONVENTIONAL HARVEST PROCESS



## Bioreactor

- Cell culture viability
- Pack cell volume
- Temperature

## Centrifugation

- Turbidity (outlet)
- Feed flow rate
- Discharge interval

## Depth filtration

- Pre-use water flush (volume, pressure, flow rate)
- Pressure drop

## Membrane filtration

- Pressure drop
- Integrity test

## Harvest Vessel

- In process testing (e.g. Titer, Bioburden, endotoxin, LDH assay, TEM)
- Yield

A blue-tinted microscopic image showing a dense network of interconnected, textured, spherical and elongated structures, likely representing cell culture or biological tissue. The structures are arranged in a somewhat vertical, branching pattern, with some larger, more rounded nodes and thinner, connecting filaments. The overall appearance is that of a complex, porous biological scaffold.

# NEW HARVEST TECHNOLOGIES FOR HIGH CELL DENSITY CELL CULTURE PROCESS

# HARVEST TECHNOLOGIES FOR HIGH DENSITY CELL CULTURE PROCESS

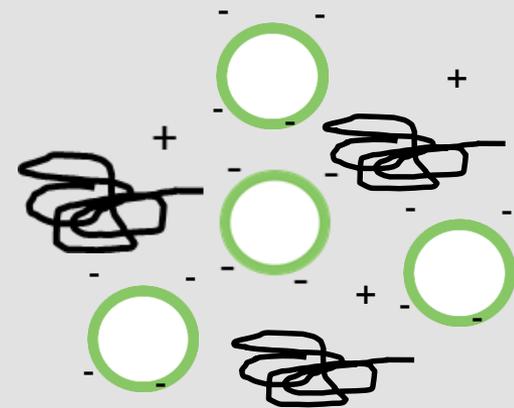
- Latest trend of cell culture processes with cell density of  $>10$  million cells and solid content of  $>15\%$
- Conventional platform process is limited by centrifuge bowl capacity and depth filter surface area, which impact on harvest yield and production cost
- Alternative technologies:
  - Accelerated sedimentation (Flocculation)
  - Tangential Flow (TF)- Microfiltration (MF)
  - Alluvial filtration
  - Acoustic Wave Separator

**INDUSTRIAL TREND IS MOVING TOWARDS HIGH  
DENSITY CELL CULTURE PROCESS**

# ACCELERATED SEDIMENTATION (FLOCCULATION)

- Particulates in CHO culture process are typically  $<10\ \mu\text{m}$ , and can be induced to form flocs or precipitates sizing between 20 and 100  $\mu\text{m}$ .
- Methods:
  - Cationic polymers (pDADMAC, PEI, Chitosan)
  - Non ionic polymers (PEG, Dextran)
  - Acid titration (to pH ~5)
  - Salt addition, for e.g  $(\text{NH}_4)_2\text{SO}_4$ ,  $\text{K}_2\text{SO}_4$ ,  $\text{KH}_2\text{PO}_4$

*Crosslinking of negatively charged  
Cells and cationic polymers  
through ionic interaction*



## Sources:

Sarah Le Merdy, *Bioprocessing journal*, Oct, 2015

*J Biotechnol*, 2007; 128(4): 813–23.

*Biotechnol Bioeng*, 2013; 110(11): 2928–37.

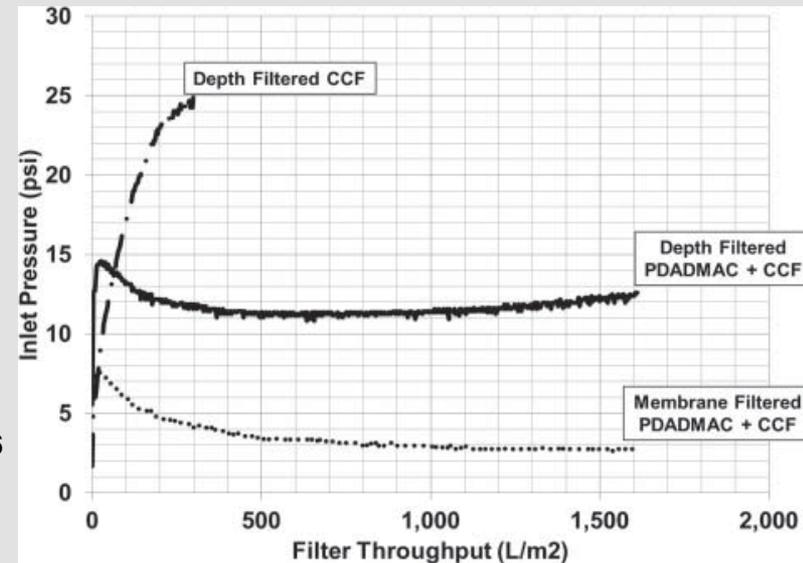
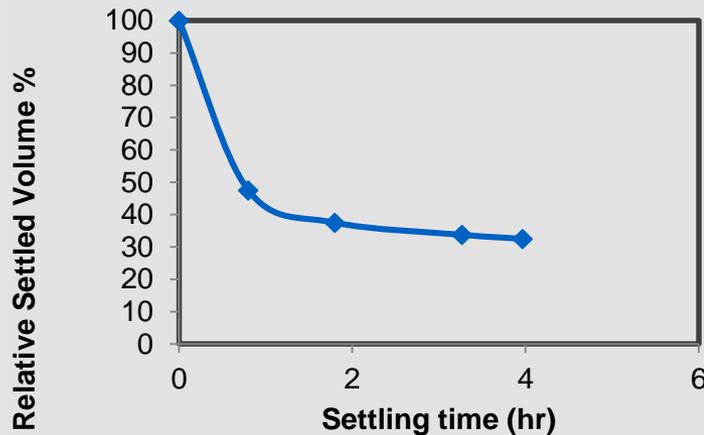
*mAbs*, 2015; 7(2): 413–27

*Biotechnol Bioeng*, 2011; 108(1): 50–8.

# FLOCCULATION USING CATIONIC POLYMER PDADMAC

McNerney et al., (2015) PDADMAC flocculation of Chinese hamster ovary cells:, mAbs, 7:2, 413-427

## Time Based Settling

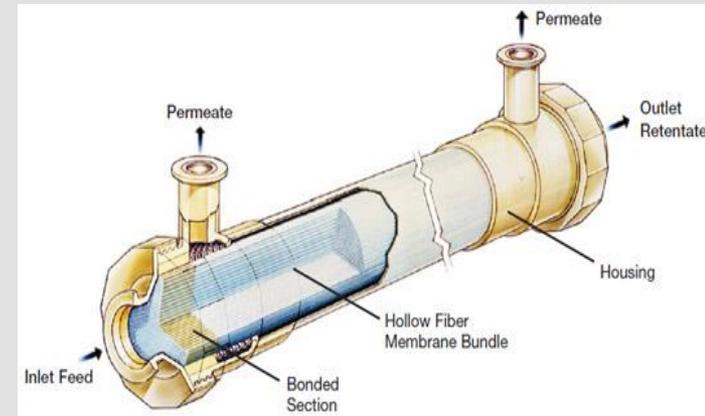
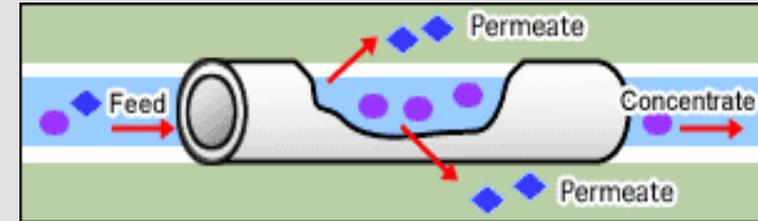
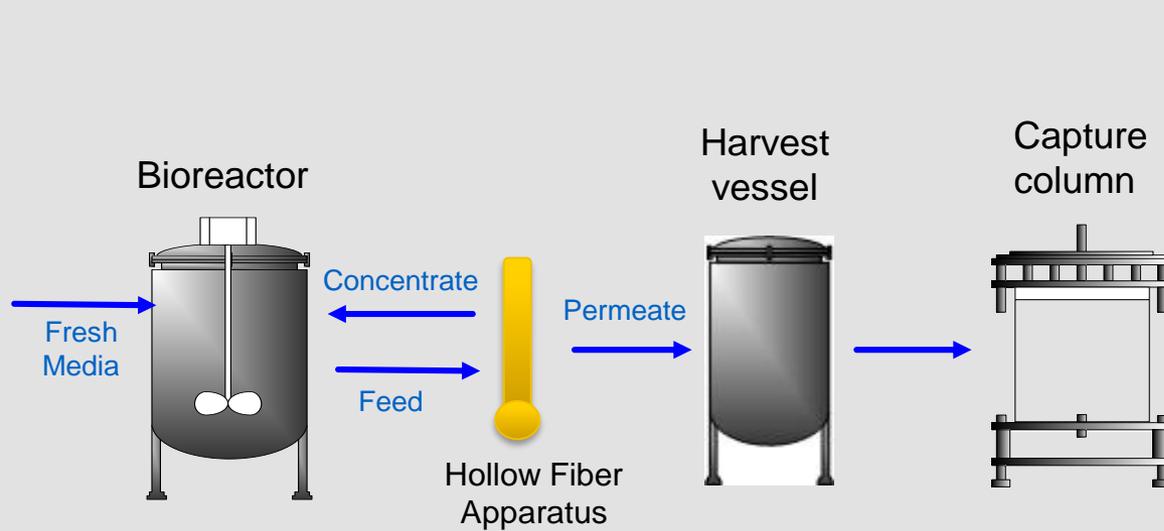


The clarity of the flocculated supernatant is comparable to centrifugation followed by depth filtration and membrane filtration

# KEY CONSIDERATION FOR FLOCCULATION METHOD

- **Evaluating impact of additives on product quality attributes**
- **Impact on downstream processing**
- **Demonstrating the downstream process clearance for additives**

# TANGENTIAL FLOW (TF)-MICROFILTRATION (MF) HARVEST SYSTEM



Sieving Coefficient

$$(S_o) = \frac{\text{Product Concentration in Permeate}}{\text{Product Concentration in Feed}}$$

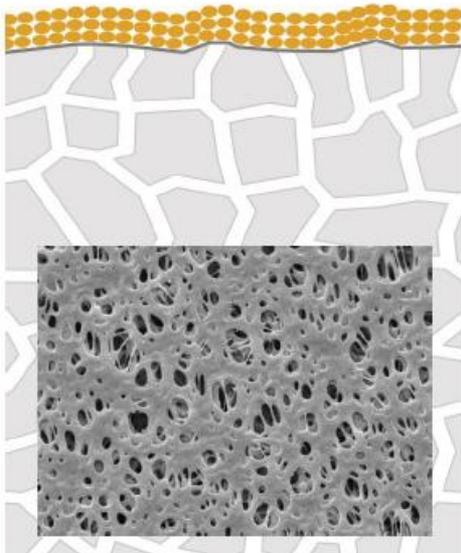
Product is small to permeate through the membrane, while cell and larger debris are retained in the concentrate line returning to bioreactor

# KEY CONSIDERATION FOR TF-MF SYSTEM

- **Filter dimension & surface area**
- **Filter pore size**
- **Cross flow rate → cell damage (shear), sweeping action**
- **Transmembrane pressure**
- **Permeate flux**

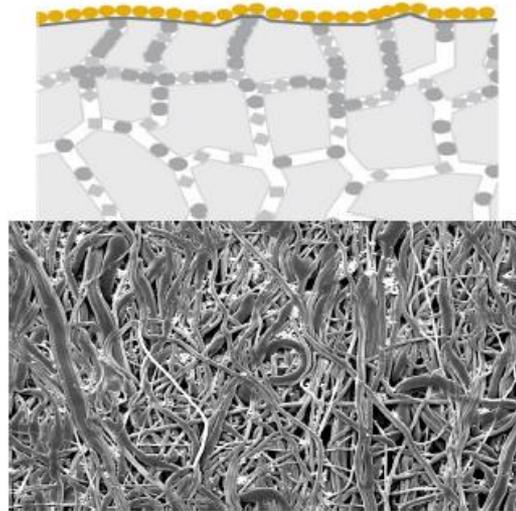
# ALLUVIAL FILTRATION

## Different Filtration Functional Principles



Surface filtration

*Membrane Filters*



Depth filtration

*Depth Filter Sheets / Modules*



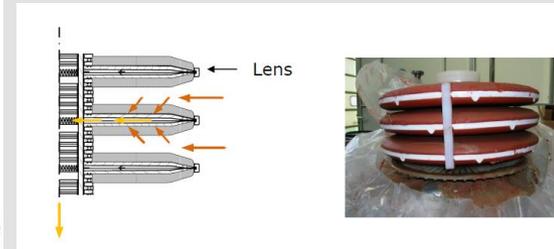
Alluvial filtration

*Body Feed / Pre-Coat Filtration*

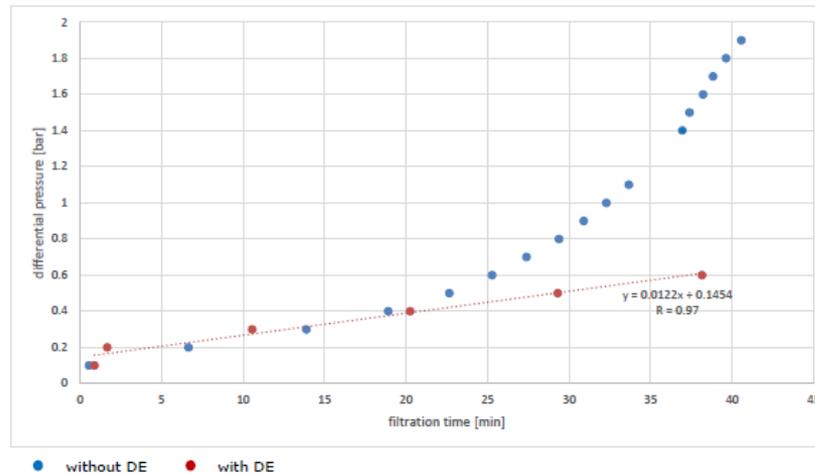
# ALLUVIAL FILTRATION

Mix the cell culture solution with filter aid material (DE)

Filtration through a specialized design filter

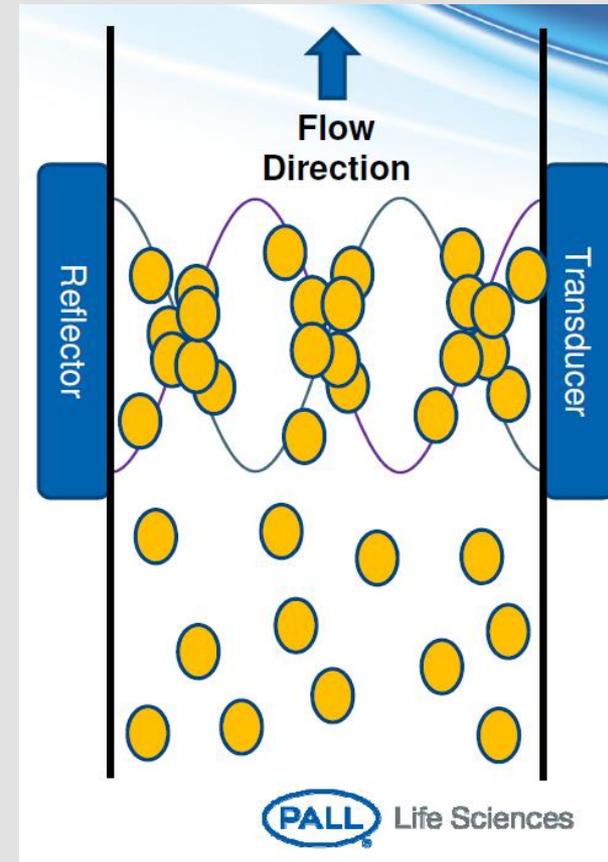


Depth Filtration vs. Alluvial Filtration / Clogging Curve



# CADENCE<sup>®</sup> ACOUSTIC SEPARATION (CAS<sup>®</sup>)

- Acoustic wave separation (AWS) technology involves the use of low frequency acoustic forces to generate a 3-dimensional (3D) standing wave across a flow channel.
- Cells are trapped by the acoustic forces, while small proteins will flow through.



# ACKNOWLEDGMENT

**Mahsa Rohani**

**Amgen Pre-Pivotal & Pivotal Process Development**